

Mr. Keith Crittenden
United Dental Resources
P.O. Box 333
Crete, IL 60417

March 11, 2008

Dear Keith,

This final report is for DNA analytical work performed on the two samples delivered by DHL to our laboratory at 1335 29 February. The analyses will be charged to the credit card number provided.

We will mail you this report and complete results.

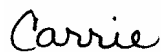
Original samples and all subcultures associated with this report will be retained for 9 days after today's date. To make arrangements to archive samples and subcultures longer than 9 days, please call.

As always I encourage you to call me at (866) 709-6600, ext. 26 with any questions you may have.

As part of our dedication to continuous improvement, we encourage you to share any suggestions or feedback you might have that would enable us to serve you better. Please send your suggestions to susan@microcheck.com or call Susan Sinclair at 866-709-6600, ext. 23.

Thank you for this opportunity to work with you.

Sincerely,



Carrie D. Pontbriand
Laboratory Manager

CDP/mgs

METHOD: Each of the dental implants was aseptically transferred from its packaging into sterile 120 milliliter (mL) plastic jars with screw top lids. 10 mL of sterile phosphate buffered water with Tween 80 (PBWT) were aseptically added to the implant in each jar. The jars were placed on a reciprocal shaker and shaken at 175 RPM for 15 minutes. Following shaking, 10-fold serial dilutions were done to isolate and enumerate any microbes from the implants. 0.1 mL from the 10 mL of PBWT in which the implant was immersed was aseptically pipeted into a tube of 0.9 mL sterile PBWT. The tube was vortexed and 0.1 mL was transferred to a second serial dilution tube containing 0.9 mL sterile PBWT. The process was repeated through a third and a fourth serial dilution tubes with the final 0.1 mL thrown away. The 0.9 mL aliquots from each of the four serial dilution tubes were poured onto separate plates of tryptic soy agar (TSA), which is a general-purpose medium on which bacteria, actinomycetes, yeasts, and filamentous fungi will grow. 0.9 mL of the original immersion PBWT aliquots were also added to TSA plates. Following incubation at 28°C the plates were examined and the apparently different colony-forming types of microbes enumerated. The colony-forming units (CFU) are expressed per implant. The individual bacterial colonies from the TSA plates were identified by their DNA.

RESULTS: Tabulated below are the LINE NUMBER (LINE #), the identification label assigned to the apparently different colony-forming type of microorganism (COLONY #), the colony-forming units per implant (CFU / IMP), the identification of the microorganism (MICROORGANISM), the percent match for the comparison (MATCH %), the type of microorganism (TYPE), the confirmation test, if necessary (CONFIRM TEST), and the lab comment (LAB COMMENTS), if appropriate.

D-1 HAN, Sample 1

LINE #	COLONY #	CFU/IMP	MICROORGANISM	MATCH %	TYPE	CONFIRM TEST	LAB COMMENT
1	1	11	<i>Bacillus simplex</i>	99.94	B		
2	2	111	<i>Staphylococcus epidermidis</i>	99.97	B		
TOTAL CFU/ML		122					

ND-2 JH, Sample 2

LINE #	COLONY #	CFU/IMP	MICROORGANISM	MATCH %	TYPE	CONFIRM TEST	LAB COMMENT
1			NO APPARENT MICROBIAL GROWTH				
TOTAL CFU/ML		0					

Results represent only the sample(s) as received. All analytical data and reports are client confidential and available only to the client. Authorization for publication of excerpts, statements, or conclusions regarding our reports is reserved pending written approval from Microcheck, Inc.

Key to Symbols and Abbreviations in the Microcheck Results Table

DNA ANALYSIS Automated 16S and LSU D2 gene sequencing for identification of aerobic and anaerobic bacteria, actinomycetes, yeast, and fungi.

* An asterisk next to an organism name indicates a GenBank search result

TYPE Microorganism TYPE

AC actinomycete

AN anaerobic bacterium

B aerobic bacterium

F fungus

() Parentheses () around an entry in the TYPE column indicate that the isolate was a different type than what the organism was submitted as by the client.

FAN facultative anaerobe

M mycobacterium

TH thermophilic bacterium

Y yeast

CONFIRM TEST CONFIRMATION TESTING is done on an isolate to confirm submission type or as requested by the client.

CONFIRM TEST RESULTS **GPR** Gram positive rods

GNR Gram negative rods

GVR Gram variable rods

coag⁺ coagulase positive

ox⁺ oxidase positive

aryl⁺ arylsulfatase positive

API 20E Metabolic characterization that is done for members of the Family Enterobacteriaceae.

GPC Gram positive cocci

GNC Gram negative cocci

coag⁻ coagulase negative

ox⁻ oxidase negative

aryl⁻ arylsulfatase negative

We encourage you to call one of our DNA Analysts with any questions you may have:

Samantha Calderon – 866/709-6600 @ ext.61

Amanda Kitchen – 866/709-6600 @ ext. 58